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Congruence and Conflict in the Higher-Level Phylogenetics of Squamate Reptiles: An Expanded Phylogenomic Perspective

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Abstract.—Genome-scale data have the potential to clarify phylogenetic relationships across the tree of life but have also revealed extensive gene tree conflict. This seeming paradox, whereby larger data sets both increase statistical confidence and uncover significant discordance, suggests that understanding sources of conflict is important for accurate reconstruction of evolutionary history. We explore this paradox in squamate reptiles, the vertebrate clade comprising lizards, snakes, and amphisbaenians. We collected an average of 5103 loci for 91 species of squamates that span higher-level diversity within the clade, which we augmented with publicly available sequences for an additional 17 taxa. Using a locus-by-locus approach, we evaluated support for alternative topologies at 17 contentious nodes in the phylogeny. We identified shared properties of conflicting loci, finding that rate and compositional heterogeneity drives discordance between gene trees and species tree and that conflicting loci rarely overlap across contentious nodes. Finally, by comparing our tests of nodal conflict to previous phylogenomic studies, we confidently resolve 9 of the 17 problematic nodes. We suggest this locus-by-locus and node-by-node approach can build consensus on which topological resolutions remain uncertain in phylogenomic studies of other contentious groups. [Anchored hybrid enrichment (AHE); gene tree conflict; molecular evolution; phylogenomic concordance; target capture; ultraconserved elements (UCE).]

Phylogenomic analyses face several major challenges. Because large data sets are used to generate these trees, many nodes in a tree often have strong statistical support (Rokas and Carroll 2006), whether measured by bootstrap or posterior probability metrics. However, this support is somewhat illusory, because alternative data sets and inference methods can yield strongly discordant results. Notable examples include the placement of ctenophores within animals (Pisani et al. 2015; Whelan et al. 2015, 2017) and relationships among bird families (Jarvis et al. 2014; Prum et al. 2015). In both cases, phylogenies were inferred with millions of sites, and most nodes in a given analysis were strongly statistically supported. Yet, some of these strongly supported nodes conflict with each other across data sets and analytical methods, suggesting that these estimates of statistical support might be inflated for some nodes (Cummings et al. 2003; Jeffroy et al. 2006). In addition—and somewhat paradoxically—phylogenomic data sets sometimes fail to provide additional resolution for some contentious nodes, despite massive amounts of data (Philippe et al. 2011).

To address these challenges, we can instead interrogate support for alternative phylogenetic hypotheses using a locus-by-locus approach (Brown and Thomson 2016; Arcila et al. 2017; Shen et al. 2017; Walker et al. 2018; Smith et al. 2020). Given the assumption of a single underlying species tree, this approach explicitly measures levels of conflict among gene trees and attempts to determine its potential causes. Researchers can then filter loci or use more sophisticated analytical methods (i.e., modeling introgression across tips, Wen et al. 2018) to better resolve nodes with high levels of conflict.

Conflict among gene trees can result from both biological processes and methodological issues. With respect to biology, certain evolutionary histories can increase gene tree conflict, including introgression among lineages, large or structured ancestral populations, and periods of rapid speciation (Maddison 1997; Degnan and Rosenberg 2006; Edwards 2009). Gene tree conflict can also arise if gene trees were estimated incorrectly due to methodological issues such as undetected paralogy, model violation, or low information content. Identifying and removing sources of gene tree estimation error can generate better-resolved phylogenies (Jeffroy et al. 2006; Salichos and Rokas 2013; Doyle et al. 2015). However, such filtering approaches cannot ameliorate gene tree incongruence that results from biological processes (but see Knowles et al. 2018). Instead, we must evaluate what these conflicts tell us about our confidence in a given node as well as the processes that have led to conflict in the first place.

Here, we apply a locus-by-locus approach to understand gene tree conflict in Squamata, the vertebrate clade comprising lizards, snakes, and

amphisbaenians. This clade includes over 10 000 species and exhibits striking instances of evolutionary convergence, with multiple independent origins of viviparity, parthenogenesis, limblessness, sex chromosomes, and venom production (Fry et al. 2006; Brandley et al. 2008; Kearney et al. 2009; Pyron and Burbrink 2014; Gamble et al. 2015; Uetz and Stylianou 2018). This group has been subject to three recent, wide-ranging phylogenomic studies (Streicher and Wiens 2016, 2017; Burbrink et al. 2020), all of which clarified key relationships among clades and identified topological relationships that remain uncertain. Building on these studies, we provide a consensus view on higher-level squamate phylogenetics by assessing conflict and congruence across thousands of independent loci, conducting targeted tests of support across high-conflict nodes, and identifying the shared properties of conflicting loci. In doing so, we show how our locus-by-locus and node-by-node approach can help focus attention on which phylogenetic relationships remain uncertain.

Methods

Sampling, Data Acquisition, and Data Processing

We used both newly collected and previously published genome-wide sequence data to infer a family-level phylogeny for squamate reptiles. We sequenced 92 target samples, prioritizing samples that were linked to vouchered museum specimens. We addressed key gaps in our phylogenetic sampling by further including 17 samples from previously published phylogenomic studies (Leaché et al. 2015; Streicher et al. 2016; Streicher and Wiens 2016; Streicher and Wiens 2017). Where possible, we downloaded the raw sequence data associated with these samples and processed them similarly to newly collected data. In total, we included 109 samples across 108 species, spanning 58 of the 67 squamate families (Supplementary Table S1 available on Dryad at http://dx.doi.org/10.5061/dryad.6p58d0k). Most families are represented by two species that span the phylogenetic breadth of the family. Our taxonomy follows Uetz and Stylianou (2018).

We used a target capture approach to sequence 5462 phylogenomic loci per newly collected sample (SqCL marker set; Singhal et al. 2017). This marker set consists of three loci types, all commonly used in vertebrate phylogenomics: 372 anchored hybrid enrichment loci (AHE; Lemmon et al. 2012), 5052 ultraconserved elements (UCE; Faircloth et al. 2012b), and 38 single-copy nuclear genes (Wiens et al. 2012). The AHE and nuclear genes are conserved exons, whereas UCEs are nonexonic conserved loci. To generate these data, we first extracted DNA from either tail or liver tissue using a high-salt or phenol-chloroform DNA extraction (Aljanabi and Martinez 1997). Following Illumina protocols, the commercial services Rapid Genomics (Gainesville, FL, USA) and Arbor Biosciences (Ann Arbor, MI, USA) then prepared dual-barcoded genomic libraries from ~1.0 ng of sheared DNA. Libraries were pooled in sets of eight; pooled libraries were then used as template for standard capture reactions following the MyBaits v3 Protocol (Arbor Biosciences; Ann Arbor, MI, USA). Following capture, libraries were pooled further and 100 libraries were sequenced per one lane of 125PE reads with the Illumina HiSeq 4000 at the University of Michigan Sequencing Core (Ann Arbor, MI, USA) and at HudsonAlpha (Huntsville, AL, USA).

We processed sequenced reads as follows: full details are available at Singhal et al. (2017). Following demultiplexing, we removed adaptor sequence using Trimmomatic v0.36 and merged overlapping reads with PEAR v0.9.6 (Zhang et al. 2013; Bolger et al. 2014). We used Trinity v2.3.2 to assemble reads and blat v36x1 to annotate assemblies (Kent 2002; Grabherr et al. 2011). To call variants per individual, we aligned trimmed reads using bwa v0.7.17 and called genotypes using GATK v3.4 (Li 2013; Van der Auwera et al. 2013). For use as outgroups, we used BLAST v2.2.29 and samtools v1.3 to extract our target loci from the human (hg38), chicken (galGal2), turtle (chrPic1), zebra finch (taeGut2), and alligator (allMis1) reference genomes (Altschul et al. 1997; Li et al. 2009).

Phylogenetic Inference

We inferred a phylogeny across species using both a coalescent-based approach (ASTRAL-III v5.5.9; Zhang et al. 2018) and concatenated approach (ExaML v3.0.19; Kozlov et al. 2015). First, we generated locus-specific alignments using mafft v7.294 (Katoh and Standley 2011). We removed any alignments that sampled <5% of individuals and then trimmed the remaining alignments to remove any individual sequences that were <300 bp and any sites that were >70% missing.

To generate a coalescent-based tree, we used RAxML v8.2.8 under the rapid hill-climbing algorithm to infer a gene tree for each locus under the GTRGAMMA model (Stamatakis 2014). To evaluate support for each gene tree, we calculated Shimodaira–Hasegawa (SH)-like values per node. We then collapsed all gene tree nodes with <10 SH-like support, resulting in an average of 9% of nodes collapsed. We used ASTRAL-III to infer a phylogeny across these gene trees.

To infer a concatenated phylogeny, we used ExaML under the CAT model. We generated 100 bootstraps by randomly subsampling 5% of the loci in the original alignment and then inferring topology with ExaML. Because bootstrapping values were uniformly high even with a small subsample and because this subsampling strategy was computationally efficient, we did not explore alternative subsampling strategies.

We then inferred both a concatenated and coalescent phylogeny using an AHE-only or UCE-only alignment, because marker type has been shown to affect phylogenetic inference (Jarvis et al. 2014; Reddy et al. 2017). We did not analyze an alignment of traditional

phylogenetic genes only due to its small sample size. Then, we identified nodes that differed among inferred trees using phyparts v0.0.1 (Smith et al. 2015). phyparts identifies concordant nodes as those that share the same set of descendants; all other nodes are discordant.

Finally, a major source of gene tree conflict can be topologies that fall into the anomaly zone, the parameter space in which gene trees are more likely to be discordant with the species tree than concordant (Degnan and Rosenberg 2006). Using scripts provided by Linkem et al. (2016), we calculated the limit of the anomaly zone for each pair of parent–child internodes (equation 4 in Degnan and Rosenberg 2006). If the descendant internal branch is shorter than the limit, this branch falls into the anomaly zone. We calculated internal branch lengths in coalescent units based on the ASTRAL-III tree.

Testing Phylogenetic Conflicts

We identified uncertain nodes in the family-level phylogeny for subsequent interrogation using several approaches. First, we identified nodes that have been resolved inconsistently across different studies (Wiens et al. 2012; Pyron et al. 2013; Streicher and Wiens 2016). In addition, we considered nodes that have been historically contentious, such as the placement of Iguania (as summarized in Losos et al. 2012). Second, we identified nodes that conflicted across the phylogenies inferred in this study (Fig. 1, Supplementary Fig. S1 available on Dryad). Third, we identified common conflicting topologies across gene trees. To do so, we used the program bp to compare rooted gene trees to the concatenated phylogeny (Smith et al. 2020). For every node, bp outputs all conflicting topologies found in the gene trees, ranked by frequency. We then manually reviewed this output to both identify high-conflict nodes and their alternate topological resolutions. Through these three approaches, we selected 17 relationships for further investigation; each had two to four alternate topological resolutions (see Table 1).

We used two complementary approaches to evaluate support for alternative topological resolutions across our 17 putatively uncertain nodes. First, we measured levels of gene tree conflict using the program bp. For a given node, if the gene tree and species tree have different descendants, bp will classify the gene tree as conflicting. We measured conflict using gene trees that were outgroup rooted and for which all nodes with <80 SH-like values were collapsed. Second, we measured the difference in log-likelihoods for a given locus across all alternate topologies, as introduced by Smith et al. (2020). Per node and locus, we calculated the log-likelihood under each alternate topology by specifying these topologies as constraints in RAxML. We then collated all likelihoods across all topological resolutions and took the difference between the two largest likelihoods as D_{LNL}. D_{LNL} is thus an estimate of the extent to which a particular topological resolution is favored over the next-best topological resolution for a given locus and node. Then, per topology, we summed D_{LNL} values across the loci that best supported that topological resolution. The summed D_{LNL} thus tells us the total weight of evidence favoring the focal topology; this metric quantifies how strongly (summed D_{LNL} large) or weakly (summed D_{LNL} small) a set of loci favors a particular topology. Similar to other measures of nodal support based on likelihood (e.g., Shen et al. 2017), the D_{LNL} approach does not account for how demographic parameters affect the likelihood of a gene tree given a species tree and thus might fail in situations like the anomaly zone (Degnan and Rosenberg 2006).

Shared Properties of Conflicting Loci

The properties of a given locus affect phylogenetic inference and thus levels of gene tree conflict (Jeffroy et al. 2006). Accordingly, we calculated 14 summary statistics that characterized the loci's overall data quality and patterns of molecular evolution (Table 2). We measured levels of missing data (missingness and occupancy), informativeness (locus length, total tree length, average SH-like value, and two metrics related to phylogenetic informativeness [PI]), heterogeneity (nucleotide compositional heterogeneity, root-tip variance, and residuals of root-tip length against root-tip node depth), quality (heterozygosity, number of long branches), GC content, and saturation C value (Townsend 2007; Kück and Struck 2014). To calculate phylogenetic informativeness, we calibrated the concatenated phylogeny using treePL (Smith and O'Meara 2012) and fossil and secondary calibrations from Irisarri et al. (2017) and then estimated PI using TAPIR (Faircloth et al. 2012a).

To determine what shared properties of loci might drive conflict, we conducted five analyses. Across all these analyses, we used the D_{LNL} results to categorize loci as either conflicting or supporting. First, per metric and per putatively contentious relationship, we calculated the mean difference between loci that supported the most-preferred topology versus those that conflicted. We then generated 1000 nonparametric bootstraps and calculated the difference between supporting and conflicting loci for each of these scrambled data sets. We calculated significance as the number of bootstraps in which the absolute difference was greater than the observed difference. Second, we determined which locus-level properties might explain the level of conflict between the gene tree and the species tree. Here, we measured the level of conflict as the difference in log-likelihoods of an unconstrained gene tree versus one constrained to the concatenated species tree. Before conducting correlations, we took the residuals of all metrics and log-likelihoods against 'tree length'. Third, we correlated patterns of D_{LNL} values across all pairwise combinations of our 17 putatively contentious nodes. Fourth, we determined if the identity of conflicting loci overlap more across topological resolutions than would be expected by random chance. To calculate the percent

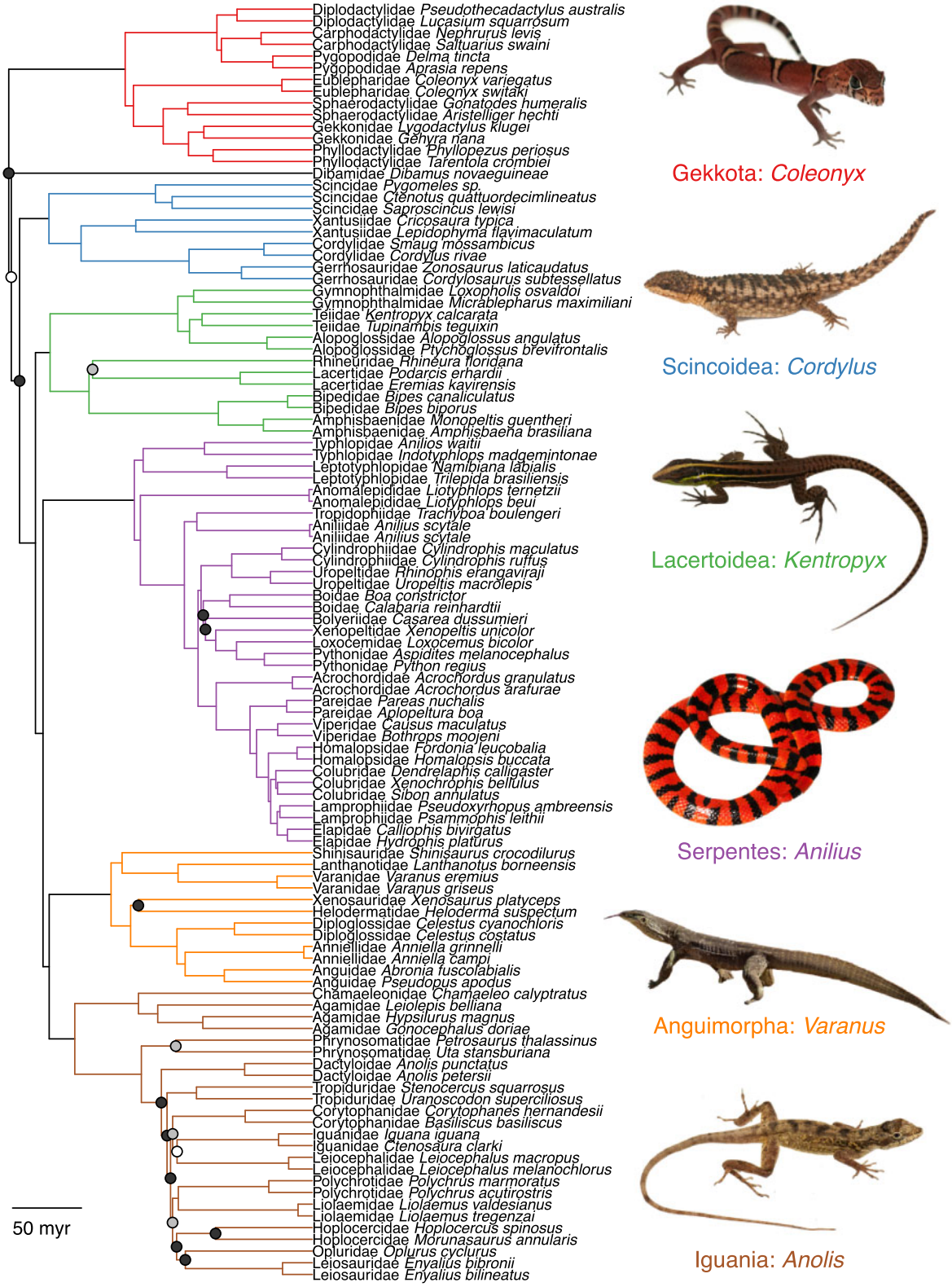

FIGURE 1. Concatenated phylogeny inferred using ExaML. Branch colors denote major squamate clades, and each clade is depicted by a representative taxon (all photographs courtesy of author TJC and Pascal Title). Nodes marked by black circles have high statistical support (bootstrap > 95) but conflict between the concatenated and coalescent-based inferred tree (Supplementary Fig. S2 available on Dryad); nodes in gray conflict and have low statistical support; nodes in white are congruent but have low statistical support. Many conflicting nodes have high statistical support.

TABLE 1. The putatively contentious relationships tested. Shown are how many alternate topologies were tested, which topology was best supported by D_{LNL} analyses (Supplementary Table S2 available on Dryad), the number and proportion of loci with $D_{LNL} > 2$ for this topology, whether this topology concords with the maximum likelihood (ML) tree (Fig. 1) and the coalescent-based tree (Supplementary Fig. S2 available on Dryad), whether this topology was resolved strongly, and if an unbiased subset of loci supported this topology (Supplementary Table S3 available on Dryad). Numbering of relationships follows Figures 2 and 3.

Uncertain relationship	No. of tested	Topology best supported by D_{LNL} analysis tested	$D_{LNL} >$ 2 loci	Matches ML tree	Matches coalescent tree	Strong resolution	Loci unbiased
1. Position of Anniellidae	3	Sister to Anguidae	857 (44%)	True	True	True	True
2. Position of Anomalepididae	3	Sister to Leptotyphlopidae and Typhlopidae	1116 (45%)	False	False	True	True
3. Position of Bolyeridae	2	Sister to Boidae	749 (48%)	False	False	False	True
4. Position of Cylindrophiidae and Uropeltidae	3	Sister to Bolyeriidae, Boidae, Pythonidae, Loxocemidae and Xenopeltidae	797 (43%)	True	False	False	True
5. Position of Dibamidae	3	Sister to all other squamates	468 (48%)	False	False	True	False
6. Position of Diplodactylidae	3	Sister to Carphodactylidae and Pygopodidae	721 (40%)	True	True	False	True
7. Position of Eublepharidae	2	Sister to Carphodactylidae, Pygopodidae and Diplodactylidae	1241 (53%)	False	False	False	True
8. Position of Gymnophthalmidae	2	Sister to Teiidae and Alopoglossidae	1290 (46%)	True	True	True	False
9. Position of Homalopsidae	4	Sister to Elapidae, Lamprophiidae, and Colubridae	1185 (50%)	True	True	True	True
10. Position of Iguania	2	Sister to Anguimorpha	4193 (88%)	True	True	True	True
11. Position of Lanthanotidae	2	Sister to Varanidae	1753 (65%)	True	True	True	True
12. Position of Rhineuridae	2	Sister to Bipedidae, Amphisbaenidae, and Lacertidae	518 (49%)	False	True	False	True
13. Position of Scincoidea	2	Sister to Episquamata	4645 (96%)	True	True	True	True
14. Position of Serpentes	2	Sister to Anguimorpha and Iguania	1784 (64%)	True	True	False	True
15. Position of Typhlopidae	3	Sister to Leptotyphlopidae	1882 (74%)	True	True	True	True
16. Position of Xenosauridae	3	Sister to Anniellidae, Anguidae, Diploglossidae, and Helodermatidae	444 (32%)	False	False	False	True
17. Relationship between Gekkota and Scincoidea	2	Gekkota sister to Scincoidea and all other squamates	1779 (59%)	True	True	True	True

overlap expected under random, we scrambled the identity of conflicting versus supporting loci for each comparison, keeping proportions constant, and then measured percent overlap across 100 bootstraps.

Finally, fifth, patterns of molecular evolution can vary across locus types. For example, UCEs contain a central conserved region and more quickly evolving flanking regions, whereas AHE exons exhibit modest levels of conservation across their entire region (Faircloth et al. 2012b; Lemmon et al. 2012; Singhal et al. 2017). To determine if locus type might affect our phylogenetic inference, we compared our locus-level metrics across all three locus types and repeated the D_{LNL} analyses for both AHE- and UCE-only data sets. We did not conduct D_{LNL} analyses with traditional phylogenetic genes because of the small sample size.

Data Analysis and Visualization

All code used to process genomic data and analyze data are available at https://github.com/singhal/conflict_analysis. We used python v3, R v3.3.3, ape, phangorn, phytools, and cowplot to process and visualize these data (Paradis et al. 2004; Schliep 2010; Revell 2012; Wilke 2016).

RESULTS

Phylogenetic Inference

Our target capture approach was highly effective; we collected an average of 4.5 Mb of sequence across 5103 loci across our 92 individuals (Supplementary Table S1 available on Dryad). Per locus, average completeness across individuals was 92%. Our newly generated data were of higher quality—higher coverage (mean 80×) and longer loci (mean 880 bp)—than previously published data, likely because of greater high sequencing effort (Supplementary Fig. S3 available on Dryad).

Using these data, we inferred both coalescent-based and concatenated trees. The two trees were largely concordant but differed at several nodes, particularly with respect to family-level relationships within Iguania (Fig. 1). Given that the two trees are fairly similar and mainly disagree at known discordant nodes, we focus further analyses and discussion on the concatenated phylogeny.

TABLE 2. A summary of the fourteen data quality and molecular evolution metrics measured for each locus. We indicate how we expect the metric to measure for "high quality" loci.

Metric	Brief description	Rationale	High quality
Branch outliers	The number of terminal branches >5× than the mean branch length	Long branch lengths can indicate poor alignment quality or mistaken ortholog identification	Lower number of branch outliers
Compositional heterogeneity RCFV	A measure of nucleotide compositional bias (Kück and Struck 2014)	Compositional heterogeneity can compromise accuracy of gene tree inference	Less heterogeneity
GC	Average proportion of sequence that is GC	High GC levels can indicate higher recombination rates, which can reduce accuracy of gene tree inference	Lower GC
Heterozygosity	Average proportion of heterozygous sites	High heterozygosity can indicate collapsed paralogs	Lower heterozygosity
Length	Average length of alignment used to infer gene tree	Long loci are more likely subject to recombination; shorter loci contain less information	Longer loci (though ambiguous)
Maximum phylogenetic informativeness (PI)	The maximum value of PI for the locus	Suggests the utility of the gene for resolving different node depths	Not clear
Maximum PI time period	In what time period the locus achieved its maximum PI	Suggests the utility of the gene for resolving different node depths	Not clear
Mean residuals	Mean residual after regressing root-tip length against root-tip node depth	High summed residuals can indicate high rate heterogeneity	Lower residuals
Missing	Average amount of missing sequence in each alignment	High levels of missing data can lead to inaccurate gene tree inference	Less missing data
Occupancy	Percent of targeted individuals included in alignment	Missing data can affect gene tree inference	Greater occupancy
Root-tip variance	The variance in branch lengths for a rooted gene tree	High root-tip variance can indicate high heterogeneity	Less variance
Saturation C value	A measure of saturation that compares the standard deviations of the transition-transversion ratio and uncorrected p-distances (Kück and Struck 2014)	High saturation can compromise accuracy of gene tree inference	Less saturation
Tree length	Total tree length	Longer trees suggest more informative sites but can also result from alignment errors	Longer trees (but ambiguous)

The concatenated phylogeny was largely concordant with previous squamate phylogenies, whether these phylogenies were inferred with a few loci or with phylogenomic data sets (Wiens et al. 2012; Streicher and Wiens 2017; Burbrink et al. 2020). However, some inferred relationships differed. For example, in the concatenated topology, Dibamidae is sister to all nongecko squamates (as in Townsend et al. 2004), whereas other studies have found it sister to all squamates (Pyron et al. 2013; Streicher and Wiens 2017) or sister to Gekkota (Wiens et al. 2012; Reeder et al. 2015; Burbrink et al. 2020). Other conflicts emerged by comparing phylogenies inferred using different marker sets and different analytical methods (Fig. 1 and Supplementary S1 available on Dryad). For example, the position of Eublepharidae differs in trees inferred with AHE versus UCE loci (Supplementary Fig. S1 available on Dryad), as seen in other studies (Townsend et al. 2004; Wiens et al. 2012; Pyron and Burbrink 2014; Reeder et al. 2015).

Testing Phylogenetic Conflicts

To more systematically evaluate conflict, we compared gene tree and species tree topologies to determine the number of gene trees that conflict at each node. Levels of support and conflict varied considerably both across clades and across clade depth (Fig. 2, Supplementary Fig. S2 available on Dryad). Although our within-family sampling was limited, monophyly of families was well-supported by the majority of gene trees (average support = 71%; Supplementary Fig. S2 available on Dryad). However, for relationships deeper than family-level, gene tree support averaged 40%. Conflict was particularly common among early branching relationships in Serpentes and Iguania; many of these branches fall into anomaly zones (Supplementary Fig. S4 available on Dryad). In fact, conflict was so rampant within Iguania that we could not identify alternate topological resolutions to test (see also Burbrink et al. 2020). Conflict was high even across nodes that had high statistical support as measured by bootstrap and local posterior probability (Supplementary Fig. S5 available on Dryad).

We then identified 17 putatively contentious nodes and used a summed log-likelihood approach to evaluate support for alternate topological resolutions at each node (Table 1). Most loci had very low D_{LNL} values (median $D_{LNL} = 1.66$; Fig. 3), indicating that they did not strongly distinguish amongst alternate

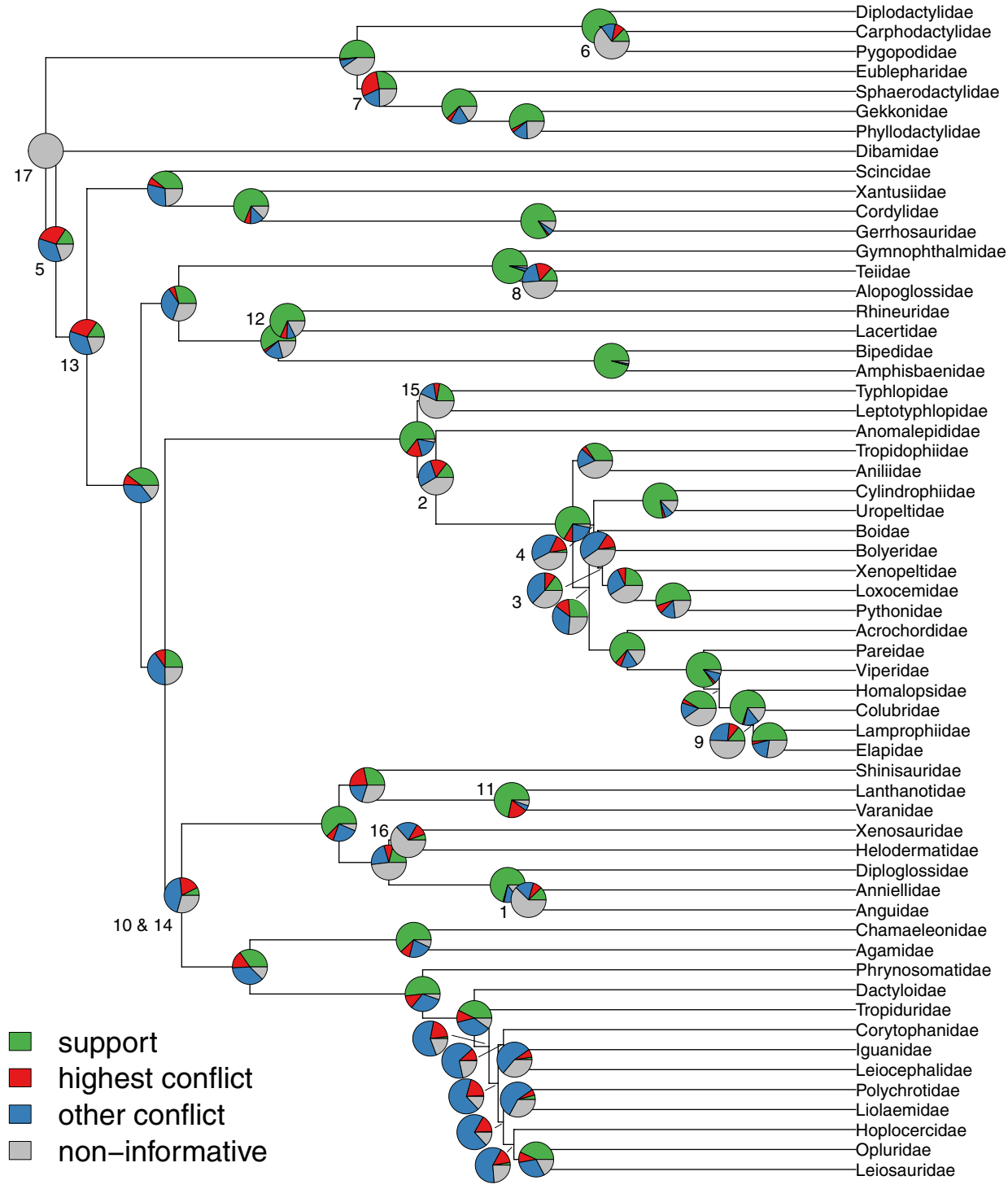

FIGURE 2. ExaML-inferred tree with levels of conflict shown at each node. Pie proportions represent the number of gene trees that either support a node, support the most common conflicting relationship, support other less common conflicting relationships, or are noninformative. Branches in gene trees with <80 SH-like support were collapsed prior to analysis. Node labels mark putatively contentious nodes; labels follow Table 1. Many nodes exhibit high levels of gene tree conflict.

topologies. Nonetheless, the summed D_{LNL} approach strongly resolved several uncertain nodes (Table 1, Supplementary Table S2 available on Dryad), including the historically contentious placement of Iguania (Losos et al. 2012, see also Burbrink et al. 2020). For 10 of the 17 nodes, comparing summed D_{LNL} across topologies provided strong support for one resolution among others (Table 1). Here, we interpret

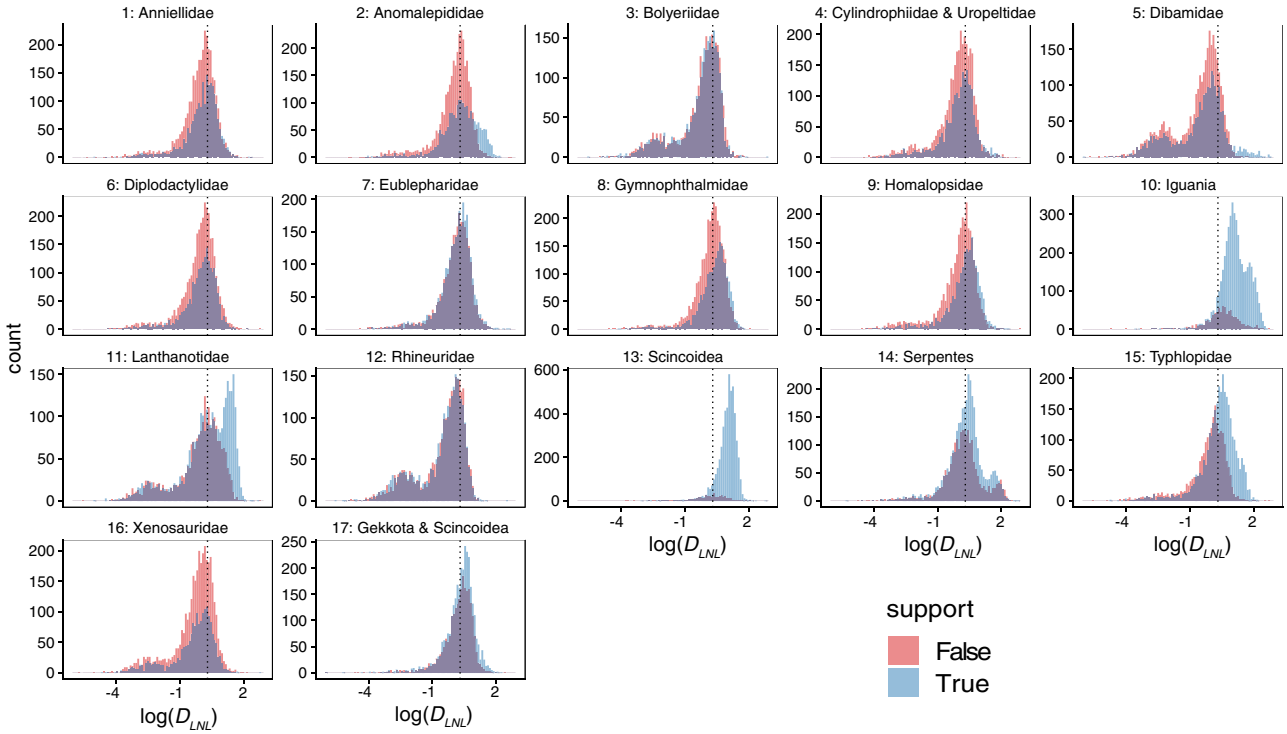

FIGURE 3. D_{LNL} values across all loci for each of the 17 putative conflicts investigated. Per locus, D_{LNL} values are measured as the difference in log-likelihoods between the two best-supported topological resolutions with respect to a focal relationship (e.g., Anniellidae). Loci are categorized by whether they support the best-supported topology (see Table 1) or not; the dotted line is where $D_{LNL} > 2$. Most loci had fairly small D_{LNL} values, suggesting they do not strongly support any given topological resolution.

a given topology as "strongly supported" when the top resolution has a summed D_{LNL} at least 50% greater than the next-best topological resolution. The summed D_{LNL} approach supported the same topology found in the concatenated tree for 11 out of 17 tested nodes (Fig. 1; Table 1). Of the remaining six nodes, four of them (position of Bolyeriidae, Eublepharidae, Rhineuridae, and Xenosauridae) had fairly equivocal support across alternate topologies—that is, alternate topological resolutions had very similar summed D_{LNL} values.

Shared Properties of Conflicting Loci

We tested if locus-specific patterns of data quality and molecular evolution could possibly be driving conflict at nodes using five approaches. First, we compared how loci properties differed between loci that supported the preferred versus alternate topologies. In general, supporting versus conflicting loci were similar across most metrics, even when these differences were significant (Table 2, Supplementary Table S3 available on Dryad). In the cases where metrics differed significantly across loci supporting different topologies, typically higher quality loci—that is, loci with less missingness, less heterogeneity—supported the preferred topology (Table 2, Supplementary Table S3 available on Dryad). Exceptions included the placement of Dibamidae, Gymnophthalmidae, and Xenosauridae, in which the best-supported topology was supported by a biased subset of lower-quality loci.

Second, we calculated the correlation between locus summary statistics and the adequacy of the concatenated topology for individual loci, finding that loci with increased compositional heterogeneity and greater root-tip variance (indicative of heterotachy) showed the greatest differences in likelihood (Fig. 4).

Third, we compared patterns of D_{LNL} values across topological tests. In general, correlations in D_{LNL} values across different tests were weak; the average correlation was $r = 0.175$ (Fig. 5A). All correlations > 0.5 were between topological tests within snakes—for example, the correlation in D_{LNL} values between "position of Cylindrophiidae and Uropeltidae" and "position of Anomalepididae."

Fourth, we determined if the identity of conflicting loci overlap more across topological resolutions than would be expected by chance, finding no more or less overlap than expected under random (Fig. 5b). Together, this result and the D_{LNL}correlations suggest little consistency in which loci conflict across different nodes.

Fifth, we repeated the summed D_{LNL} tests with AHE loci only, finding patterns in agreement with the full data set at 11 of the 17 contentious nodes (Supplementary Table S4 available on Dryad). Of the remaining six, the summed D_{LNL} values across topological resolutions were similar, suggesting that the D_{LNL} test was inconclusive. Finally, the AHE markers

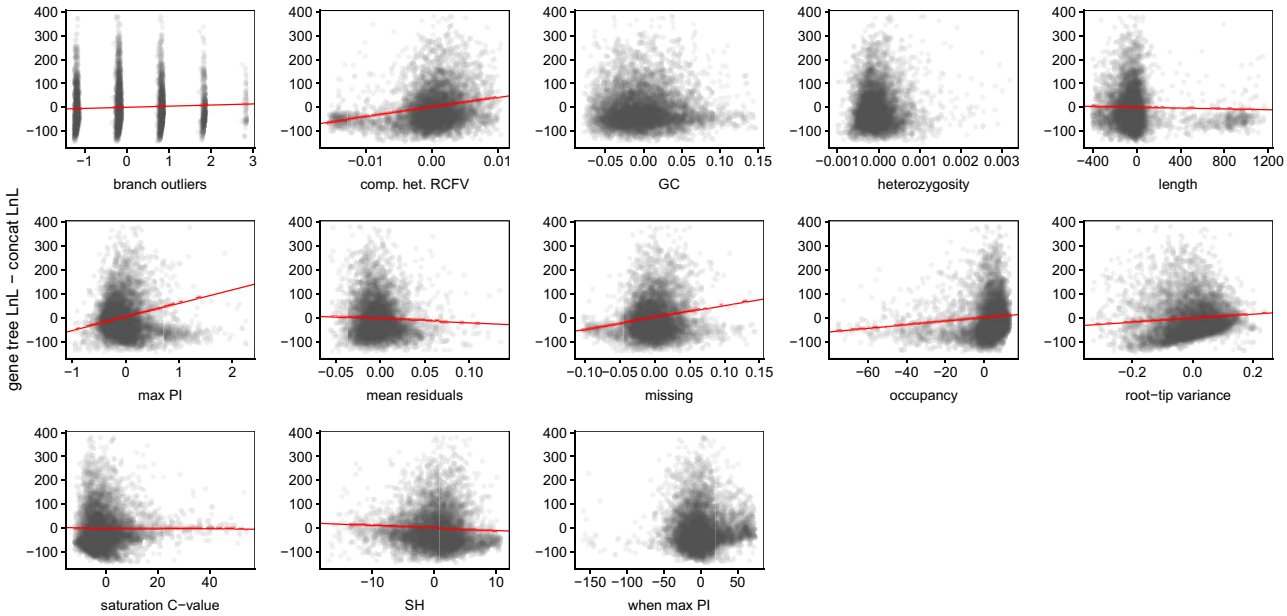

FIGURE 4. Correlation between locus summary statistics (Table 2) and the level of conflict between gene trees as species trees across marker types. Conflict level was measured as the difference in log-likelihoods of an unconstrained gene tree versus one constrained to the concatenated species tree. Larger values suggest greater conflict. Linear model fit shown for significant correlations as measured by Spearman's correlation and shown for visualization only. The strongest absolute correlations are for compositional heterogeneity (Spearman's $\rho = 0.19$; $P = 3.0\text{e-}49$) and root-tip variance (Spearman's $\rho = 0.18$; $P = 1.3\text{e-}39$). These results suggest that loci with greater compositional heterogeneity or greater rate variation across the tree are more likely to differ from the concatenated topology.

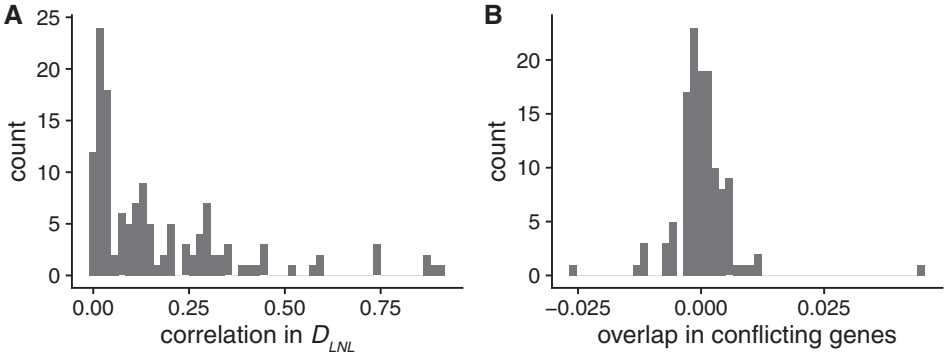

FIGURE 5. a) Correlation in locus D_{LNL} values across different topological tests. The mean correlation in D_{LNL} is $r = 0.175$; the few correlations >0.5 all stem from topological comparisons within snakes. b) Percent overlap in conflicting loci across different topological tests, shown as the mean deviation from percent overlap of 100 random bootstraps. Values >0 indicate greater overlap than expected by random. Together, these results suggest that there is little to modest consistency in which loci conflict across different nodes.

generally exhibited less conflict with the species tree, had less missing data and were more informative, and showed less evidence of heterogeneity (Fig. 6, Supplementary Fig. S6 available on Dryad).

DISCUSSION

Squamate Phylogenomics

Our 5343-locus phylogeny captures 86% of the family-level diversity in squamate reptiles and recapitulates many of the same relationships identified by studies with more taxa and fewer loci (Pyron et al. 2013; Tonini et al. 2016) and similar phylogenomic data sets (Streicher and Wiens 2017; Burbrink et al. 2020). Many of the differences between our tree and previously published trees—for example, relationships among gecko families, placement of Xenosauridae, placement of Dibamidae—have shown instability across studies that either sample different loci and taxa and/or use different analytical methods. We replicate this pattern of discordance in our study, finding topological differences across trees inferred using concatenated versus coalescent-based methods (Fig. 1), as well as for UCE versus AHE loci only (Supplementary Fig. S1 available on Dryad). Given that levels of gene tree

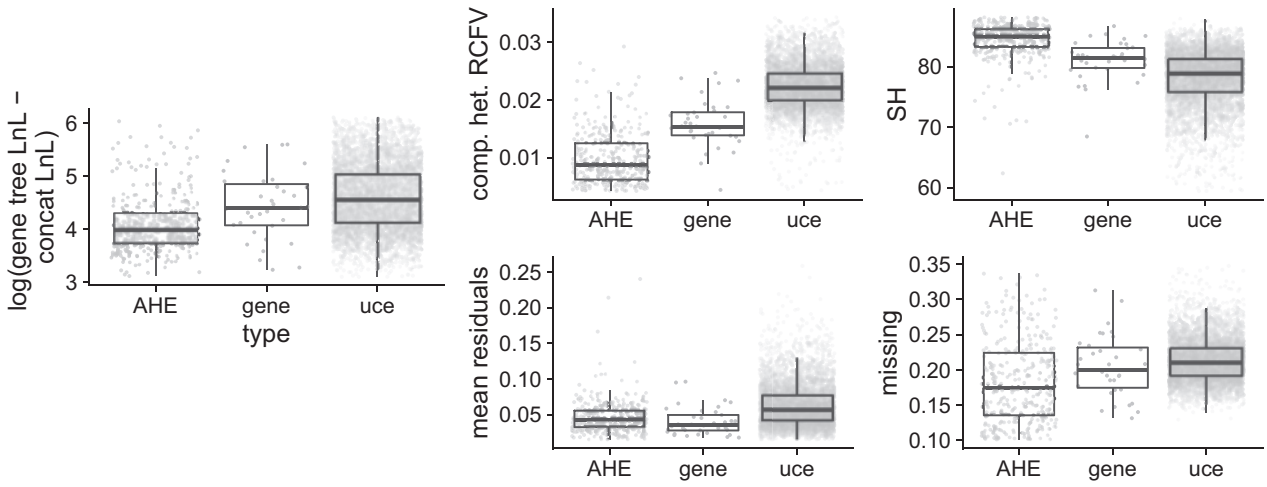

FIGURE 6. Comparative performance across the three marker types used in this study: anchored hybrid enrichment (AHE) markers, standard phylogenetic genes, and ultraconserved elements (UCEs). a) The level of conflict between gene trees as species trees across marker types. Conflict level was measured as the difference in log-likelihoods of an unconstrained gene tree versus one constrained to the concatenated species tree. These results suggest that AHE loci better fit the concatenated tree. b) Differences in locus quality metrics across marker types. In general, AHE markers showed evidence of being higher quality (i.e., they had lower levels of missingness) and more informative (i.e., trees inferred with AHE markers had higher nodal support as measured by Shimodaira–Hasegawa [SH]-like support).

conflict are high for most nodes in the phylogeny (Fig. 2, Supplementary Fig. S2 available on Dryad), this discordance across data sets and studies is perhaps unsurprising.

We explored 17 putatively contentious nodes in detail. Some of these are nodes that have low statistical support, some are nodes that have alternate topologies depending on the data set and analytical method used, and others show extensive gene tree conflict. By comparing summed D_{LNL} values across topological resolutions, we could strongly resolve 10 of these 17 nodes (Table 1). However, although the placements of Dibamidae and Gymnophthalmidae were strongly resolved, they should remain open questions. For both, support for the preferred topology is partially driven by markers with greater data missingness and more heterogeneity, and for Dibamidae, relatively few markers were sampled.

Further, 8 of these 10 strongly supported topological resolutions were also recovered in the concatenated phylogeny. The exceptions are the placements of Dibamidae and Anomalepididae. Anomalepididae, along with Leptotyphlopidae and Typhlopidae, constitutes the blind snakes, a group of fossorial snakes with reduced eyes. Most phylogenetic studies have placed Anomalepididae as either sister to all snakes or sister to all nonblind snakes (Streicher and Wiens 2016). In all inferred phylogenies (Fig. 1, Supplementary Figs S1 and S2 available on Dryad), we recover Anomalepididae as sister to all nonblind snakes, which would suggest the ancestor of all snakes likely resembled blind snakes (Bellairs and Underwood 1951). In contrast, our D_{LNL} results recover Anomalepididae as sister to other blind snakes (Table 1), as found in phylogenetic studies that consider morphological data (Hsiang et al. 2015). However, our D_{LNL} analysis based solely on AHE loci weakly supports Anomalepididae as sister to all nonblind snakes (Supplementary Table S4 available on Dryad). Supporting versus conflicting loci for Anomalepididae are similar across all measured metrics (Supplementary Table S3 available on Dryad); thus, this discrepancy between our topologies and D_{LNL} results might result from variance at some other unmeasured metric of the sampled loci (e.g., gappiness of alignment).

Comparing our results to other phylogenomic analyses (Streicher and Wiens 2016, 2017; Burbrink et al. 2020), we can build consensus on which relationships in the squamate phylogeny remain uncertain. These three studies and ours employ different sampling, similar marker sets (either AHEs or UCEs or both), and different approaches to inferring nodal support (bootstrap, local posterior probability, or locus-by-locus approaches). Thus, they can be regarded as semi-independent studies. Summarizing these studies suggests that 9 of the 17 putatively contentious nodes in Squamata have been resolved (Table 3). Most notable among these eight nodes is the placement of Iguania, which has been historically contentious (Losos et al. 2012). Further, like Burbrink et al. (2020), we find no evidence that biased loci drive the placement of Iguania (Supplementary Table S3 available on Dryad), as has been suggested in previous analyses (Gauthier et al. 2012; Koch et al. 2018). A few nodes—for example, the placement of Dibamidae, the position of Eublepharidae—remain uncertain and also have low statistical support across studies (Table 3). However, we also identify a few nodes—for example, position of Bolyeridae, position of Cylindrophiidae and Uropeltidae, position of Xenosauridae—which both have strong statistical support and conflicting topologies across previous studies. Our D_{LNL} analysis identified

TABLE 3. Summary of putatively uncertain nodes across our study and three previous phylogenomic studies in squamates (Burbrink et al. 2020; Streicher and Wiens 2016, 2017). For each relationship, we list the node, which node was best supported by our D_{LNL} analyses, which node was recovered in our maximum likelihood tree, and which node was recovered in the Burbrink et al. (2020) coalescent-based phylogeny and the Streicher and Wiens (2016, 2017) ML concatenated phylogenies, and whether or not the consensus across results suggests if this node remains uncertain. The two phylogenies inferred by Streicher and Wiens (2016, 2017) are shown in a single column because they span largely nonoverlapping parts of the squamate phylogeny. Relationships marked with an asterisk have weak support, either as measured by relative summed D_{LNL} values, local posterior probabilities, or bootstrap values.

Uncertain relationship	Topology best supported by D_{LNL} analysis	ML tree	Burbrink et al. (2020)	Streicher and Wiens (2016, 2017)	Remains uncertain
1. Position of Anniellidae	Sister to Anguidae	Sister to Anguidae	sister to Anguidae	NA	No
2. Position of Anomalepididae	Sister to Leptotyphlopidae and Typhlopidae	Sister to all nonblind Snakes	Sister to all nonblind Snakes	Sister to all nonblind snakes	Yes
3. Position of Bolyeridae	*Sister to Boidae	Sister to Pythonidae, Loxocemidae, and Xenopeltidae	*Sister to Pythonidae, Loxocemidae, and Xenopeltidae	*Sister to Pythonidae, Loxocemidae, and Xenopeltidae	Yes
4. Position of Cylindrophiidae and Uropeltidae	*Sister to Bolyeriidae, Boidae, Pythonidae, Loxocemidae, and Xenopeltidae	Sister to Bolyeriidae, Boidae, Pythonidae, Loxocemidae, and Xenopeltidae	sister to Bolyeridae, Boidae, Pythonidae, Loxocemidae, and Xenopeltidae	*sister to Acrochordidae, Bolyeridae, Boidae, Colubridae, Elapidae, Homalopsidae, Pythonidae, Lamprophiidae, Pareidae, Viperidae, Loxocemidae, and Xenopeltidae	Yes
5. Position of Dibamidae	Sister to all other squamates	*Sister to all nongeckos	*Sister to Gekkota	*Sister to all other squamates	Yes
6. Position of Diplodactylidae	*Sister to Carphodactylidae & Pygopodidae	Sister to Carphodactylidae and Pygopodidae	Sister to Carphodactylidae and Pygopodidae	*Sister to Carphodactylidae and Pygopodidae	No
7. Position of Eublepharidae	*Sister to Carphodactylidae, Pygopodidae, and Diplodactylidae	Sister to Phyllodactylidae, Gekkonidae, and Sphaerodactylidae	*Sister to all geckos	*Sister to all geckos	Yes
8. Position of Gymnophthalmidae	Sister to Teiidae and Alopoglossidae	Sister to Teiidae and Alopoglossidae	NA	NA	Yes
9. Position of Homalopsidae	Sister to Elapidae, Lamprophiidae, and Colubridae	Sister to Elapidae, Lamprophiidae, and Colubridae	Sister to Elapidae, Lamprophiidae, and Colubridae	Sister to Elapidae, Lamprophiidae, and Colubridae	No
10. Position of Iguania	Sister to Anguimorpha	Sister to Anguimorpha	*Sister to Anguimorpha	Sister to Anguimorpha	No
11. Position of Lanthanotidae	Sister to Varanidae	Sister to Varanidae	NA	Sister to Varanidae	No
12. Position of Rhineuridae	*Sister to Bipedidae, Amphisbaenidae, and Lacertidae	*Sister to Lacertidae	NA	*Sister to Lacertidae	Yes
13. Position of Scincoidea	Sister to Episquamata	Sister to Episquamata	Sister to Episquamata	Sister to Episquamata	No
14. Position of Serpentes	*Sister to Anguimorpha and Iguania	Sister to Anguimorpha and Iguania	*Sister to Anguimorpha and Iguania	Sister to Anguimorpha and Iguania	No
15. Position of Typhlopidae	Sister to Leptotyphlopidae	Sister to Leptotyphlopidae	Sister to Leptotyphlopidae	*Sister to all other snakes but Leptotyphlopidae	No
16. Position of Xenosauridae	*Sister to Anniellidae, Anguidae, Diploglossidae, and Helodermatidae	Sister to Helodermatidae	Sister to Anniellidae, Anguidae, and Diploglossidae	*Sister to Anniellidae, Anguidae, and Diploglossidae	Yes
17. Relationship between Gekkota and Scincoidea	Gekkota sister to Scincoidea and all other squamates	Gekkota sister to Scincoidea and all other squamates	Gekkota sister to Scincoidae and all other squamates	*Gekkota sister to Scincoidae and all other squamates	No

these nodes as having ambiguous support, even when traditional measures of support failed to capture this ambiguity. These results suggest the power of locus-by-locus approaches to identify contentious nodes in phylogenies. Below, we explore potential causes for this conflict at these contentious nodes.

Sources of Conflict

Biological sources of conflict.— Gene tree conflict can arise from multiple biological sources—incomplete lineage sorting, introgression, gene duplication, or varying selective or recombination regimes across loci (Maddison 1997; Degnan and Rosenberg 2006; Duchêne et al. 2018). Of these sources of conflict, incomplete lineage sorting—particularly as it arises during rapid radiation (e.g., Cloutier et al. 2019) —most likely affects our data set. Many of the internode distances within snakes and iguanids are very short (Fig. 1), which could reflect rapid radiations in these clades. Accordingly, we tested if any branches in our tree are in anomaly zones (Degnan and Rosenberg 2006). We found that relationships within Iguania and within the clade spanning Boidae to Pythonidae in Serpentes are in anomaly zones (Supplementary Fig. S4 available on Dryad). Both 'position of Bolyeridae' and 'position of Cylindrophiidae and Uropeltidae' fall within the anomaly zone in Serpentes (see Table 1), which limits our ability to interrogate these nodes using likelihood-based tests. Nonetheless, our tests of these nodes were inconclusive (Table 3). In such cases where poor resolution is driven by biological processes, phylogenetic uncertainty cannot be simply addressed through better sampling, and these relationships are likely to persist as unresolvable.

Gene tree estimation error: uninformative loci.— If loci have low information content, then some nodes in the inferred gene tree can be essentially resolved randomly. This leads to extensive gene tree conflict, although this conflict does not necessarily impact the reliability of species tree inference (Lanier et al. 2014; Blom et al. 2016). To test if uninformative loci are driving conflict, we measured locus properties that reflect information content, including SH values, tree length, locus length, and phylogenetic informativeness. Generally, we found loci with greater informativeness (greater locus length, higher SH, greater tree length) had higher concordance with our species tree (Fig. 4, see also Burbrink et al. 2020), though results across phylogenetic informativeness were mixed. In our data set, more than 68% of our loci reached their maximum phylogenetic informativeness >100 Ma (Supplementary Fig. S7 available on Dryad). Most of our loci should thus have adequate power to inform deeper relationships in squamates, such as family-level relationships within Iguania, many of which formed ~80–100 Ma. Yet, most loci exhibit only minimal differences in log-likelihoods across competing relationships (Fig. 3), suggesting these loci might be weakly informative about these deeper nodes. Indeed, on average, only 2424 of the 5354 loci sampled offered strong support for one relationship over another ($D_{LNL} > 2$). Possibly, loci with greater information content—perhaps ones that are longer or that evolve more quickly—might be more variable in their relative likelihoods across these relationships. However, perhaps because of the low correlation of loci D_{LNL} values across nodes (Fig. 5a), we found no relationship between a locus's average D_{LNL} and our measures of loci informativeness.

Gene tree estimation error: model violation.— Model violation is an important source of gene tree estimation error. We quantified several metrics of loci and their inferred trees that suggest the potential for model violation. For example, high root-tip variance might reflect rate heterogeneity across lineages, high compositional heterogeneity might reflect biased mutational process, high GC might reflect high recombination rates (Romiguier et al. 2016), and high saturation c-values might reflect multiple mutations to the same position. In our pipeline, we implemented fairly simple models of sequence and tree evolution. Particularly for UCEs—in which there is marked spatial heterogeneity in rates of evolution across the locus—these models might be too simple which could then lead to gene tree estimation error (but see Abadi et al. 2019). Such model violation might partially explain why loci with greater rate and compositional heterogeneity showed the greatest difference between unconstrained and constrained gene tree likelihoods (Fig. 4), why many of the loci supporting alternate, less-supported topologies exhibited higher rates of rate and compositional heterogeneity (Supplementary Table S3 available on Dryad), and why AHE gene trees showed better fit to the species tree than UCE gene trees (Fig. 6).

Gene tree estimation error: poor data quality.— In a phylogenomic pipeline, data quality issues can arise across multiple steps, including poor sequencing quality, misassemblies, and mistaken ortholog identification. These technical issues can result in messy alignments, which could include poorly aligned regions or regions with high missingness. The gene trees inferred from these alignments might then have inaccurate topologies (Wong et al. 2008) or have longer branch lengths, more branch outliers, or show higher levels of root-tip variance. Together, these sources of error can create gene tree conflict even if they do not necessarily impact species tree inference (Nute et al. 2018). We attempted to mitigate some of these quality issues by trimming alignments and requiring strict orthology identification. Yet, we still see evidence for variance across all these metrics of locus and tree quality (Supplementary Fig. S7 available on Dryad). In particular, loci with high levels of missingness and greater number of branch outliers exhibit bigger log-likelihood differences in unconstrained topologies

versus topologies constrained to the species tree (Fig. 4), and we found conflicting loci were more likely to have greater missingness (Supplementary Table S3 available on Dryad). Emerging tools like SpruceUp and TreeShrink (Mai and Mirarab 2018; Borowiec 2019) automatically profile alignments and inferred trees, offering a promising way to identify and remove low-quality samples and loci that can increase gene tree conflict.

Comparisons across marker types.— Other analyses have found the type of marker—for example, intron versus exon—can influence phylogenetic inference (Jarvis et al. 2014; Reddy et al. 2017). In this study, we sequenced three markers types, which are relatively similar. These markers all have relatively slow evolutionary rates (Faircloth et al. 2012b; Lemmon et al. 2012), and they almost certainly evolved under a history of purifying selection (Katzman et al. 2007). Despite these similarities, AHE markers have less missing data, exhibit less heterogeneity, and are more informative than UCE markers or genes (Fig. 6b). These locus-level properties reduce discordance between gene trees and the species tree (Fig. 5). Consequently, AHE markers show smaller differences in log-likelihoods between their unconstrained topologies and the topologies constrained to the species tree (Fig. 6a). Despite the differences in quality across marker types, an AHE-only D_{LNL} analysis returned a concordantly strong resolution for nine of the ten contentious nodes resolved strongly by the full data set (Supplementary Table S4 available on Dryad).

Phylogenomics and Phylogenetic Conflict

In many phylogenomic studies, independent analyses of the same clade often return trees that conflict with one another yet have high statistical support (e.g., Pisani et al. 2015; Whelan et al. 2015, 2017). Here, we recapitulate this finding; our inferred trees have nodes that conflict with those found in three other squamate phylogenomic studies (Table 3, Streicher and Wiens 2016, 2017; Burbrink et al. 2020). Several of these conflicting nodes have strong statistical support, but our D_{LNL} analysis identifies these nodes as remaining uncertain—thus showing the power of a locus-by-locus and node-by-node approach.

Further, although our trees conflict, assessments of which nodes are likely resolved—and which nodes remain uncertain—are robust across the locus-by-locus and node-by-node analysis we conducted and one conducted by Burbrink et al. (2020). We independently designed different studies to address the same question, using different marker and taxon sets and different gene-wise analyses to assess support and conflict. Yet, both Burbrink et al. (2020) and our study found the same pattern across the two nodes we both tested; both studies strongly supported a nested relationship for Iguania and showed uncertainty in the placement of Dibamidae. Although the number of shared comparisons is small, this concordance suggests this locus-by-locus and node-by-node approach provides better insights into levels of support for particular topological resolutions (Shen et al. 2017; Walker et al. 2018; Smith et al. 2015), relative to traditional measures that use clade posterior probabilities or bootstrap proportions (see also Supplementary Fig. S5 available on Dryad). Thus, this general approach of interrogating nodes might help build consensus across different phylogenomic studies on which nodes are resolved and which remain uncertain. Based on this consensus, future researchers could then target uncertain relationships with different locus or taxon sampling or improved analytical methods.

Other studies have argued to filter loci to ameliorate gene tree conflict (Jeffroy et al. 2006; Doyle et al. 2015; Whelan et al. 2015), specifically removing loci with low information content. Particularly for coalescent-based methods, less-informative loci tend to lead to less accurate gene trees, which could lead to inaccurate species trees (Gatesy and Springer 2014 but see Blom et al. 2016). Removing such loci often results in better-resolved species trees. Our results suggest, however, that supporting versus conflicting loci do not dramatically differ in information content (Fig. 4, Supplementary Table S3 available on Dryad), suggesting low information content might simply increase noise rather than introducing bias.

Employing a locus-by-locus approach sidesteps this debate. Instead of removing less-informative loci, we quantified how much support a given locus has for a particular topology relative to others. Most loci show only minimal differences in likelihoods across different constrained topologies (Fig. 3), which accords with a more general finding that only a small proportion of sequenced loci can drive overall phylogenetic patterns (Brown and Thomson 2016; Shen et al. 2017). Further, different loci have power to resolve nodes in different parts of the phylogenetic tree. For example, we see little correlation in D_{LNL} values across loci for different tested relationships, even across adjacent nodes or nodes with similar splitting times (Fig. 5a). Filtering loci on general informativeness risks removing loci that might inform specific relationships (Chen et al. 2015; Dornburg et al. 2019; Smith et al. 2020). Instead, the pipeline used here, where we ensure that biased loci are not driving topological resolutions, provides an alternative approach (Supplementary Table S3 available on Dyrad). Better identification and then removal of loci with poor data quality—for example, mistaken orthology assignment, chimeric assemblies—from large phylogenomic data sets could further strengthen this approach.

Finally, traditionally, downstream phylogenetic analyses such as ancestral state reconstruction have incorporated uncertainty in topologies by sampling across bootstrapped trees or a posterior distribution. But, when inferred from phylogenomic data, bootstrap trees and posterior distributions often fail to properly capture the uncertainty inherent in evolutionary

relationships (e.g., Arcila et al. 2017; Smith et al. 2020). A potential solution is to conduct comparative analyses across gene trees, particularly in cases where gene tree conflict is driven by biological processes (Hahn and Nakhleh 2016). An additional solution might be to develop new approaches for translating these alternative measures of nodal support (e.g., number of gene trees supporting a given node, summed log-likelihoods) into uncertainty metrics that can then be properly modeled in comparative analyses. As we collect larger and larger phylogenomic data sets, such advances, along with improved methods for inferring and modeling sources of conflict, will allow us to both better generate robust phylogenies and to use these phylogenies to understand the evolution of life's diversity.

Data Availability

- All scripts used in processing and analyzing the data: https://github.com/singhal/conflict_analysis
- Short-read sequencing data: BioProject PRJNA650555; SRA accessions provided in Supplementary table S1 available on Dryad
- Locus alignments and tree topologies: http://dx.doi.org/10.5061/dryad.6p58d0k.
- Many sequenced individuals are accessioned as museum specimens (Supplementary Table S1 available on Dryad)

Supplementary Material

Data available from the Dryad Digital Repository: http://dx.doi.org/10.5061/dryad.6p58d0k.

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References

Abadi S., Azouri D., Pupko T., Mayrose I. 2019. Model selection may not be a mandatory step for phylogeny reconstruction. Nat. Commun. 10:934.

Aljanabi S.M., Martinez I. 1997. Universal and rapid salt-extraction of high quality genomic DNA for PCR-based techniques. Nucleic Acids Res. 25:4692–4693.

Altschul S.F., Madden T.L., Schäffer A.A., Zhang J., Zhang Z., Miller W., Lipman D.J. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res. 25:3389–3402.

Arcila D., Ortí G., Vari R., Armbruster J.W., Stiassny M.L.J., Ko K.D., Sabaj M.H., Lundberg J., Revell L.J., Betancur-R R. 2017. Genome-wide interrogation advances resolution of recalcitrant groups in the tree of life. Nat. Ecol. Evol. 1:20.

Bellairs A. d'A, Underwood G. 1951. The origin of snakes. Biol. Rev. 26:193–237.

Blom M.P.K., Bragg J.G., Potter S., Moritz C. 2016. Accounting for uncertainty in gene tree estimation: summary-coalescent species tree inference in a challenging radiation of Australian lizards. Syst. Biol. 66:352–366.

Bolger A.M., Lohse M., Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120.

Borowiec, M., 2019. Spruceup: fast and flexible identification, visualization, and removal of outliers from large multiple sequence alignments. J. Open Source Softw. 4:1635.

Brandley M.C., Huelsenbeck J.P., Wiens J.J. 2008. Rates and patterns in the evolution of snake-like body form in squamate reptiles: evidence for repeated re-evolution of lost digits and long-term persistence of intermediate body forms. Evol. Int. J. Org. Evol. 62:2042–2064.

Brown J.M., Thomson R.C. 2016. Bayes factors unmask highly variable information content, bias, and extreme influence in phylogenomic analyses. Syst. Biol. 66:517–530.

Burbrink F.T., Grazziotin F.G., Pyron R.A., Cundall D., Donnellan S., Irish F., Keogh J.S., Kraus F., Murphy R.W., Noonan B., Raxworthy C.J., Ruane S., Lemmon A.R., Lemmon E.M., Zaher H. 2020. Interrogating genomic-scale data for Squamata (lizards, snakes, and amphisbaenians) shows no support for key traditional morphological relationships. Syst. Biol. 69:502–520.

Chen M.Y., Liang D., Zhang P., 2015. Selecting question-specific genes to reduce incongruence in phylogenomics: a case study of jawed vertebrate backbone phylogeny. Syst. Biol. 64:1104–1120.

Cloutier A., Sackton T.B., Grayson P., Clamp M., Baker A.J., Edwards S.V. 2019. Whole-genome analyses resolve the phylogeny of flightless birds (Palaeognathae) in the presence of an empirical anomaly zone. Syst. Biol. 68:937–955.

Cummings M.P., Handley S.A., Myers D.S., Reed D.L., Rokas A., Winka K. 2003. Comparing bootstrap and posterior probability values in the four-taxon case. Syst. Biol. 52:477–487.

Degnan J.H., Rosenberg N.A. 2006. Discordance of species trees with their most likely gene trees. PLoS Genet. 2:e68.

Dornburg, A., Su, Z., Townsend, J.P. 2019. Optimal rates for phylogenetic inference and experimental Design in the era of genome-scale data sets. Syst. Biol. 68:145–156.

Doyle V.P., Young R.E., Naylor G.J.P., Brown J.M. 2015. Can we identify genes with increased phylogenetic reliability? Syst. Biol. 64:824–837.

Duchêne D.A., Bragg J.G., Duchêne S., Neaves L.E., Potter S., Moritz C., Johnson R.N., Ho S.Y., Eldridge M.D. 2018. Analysis of phylogenomic tree space resolves relationships among marsupial families. Syst. Biol. 67:400–412.

Edwards S.V. 2009. Is a new and general theory of molecular systematics emerging? Evolution 63:1–19.

Faircloth B.C., Chang J., Alfaro M.E. 2012a. TAPIR enables high-throughput estimation and comparison of phylogenetic informativeness using locus-specific substitution models. arXiv Prepr. arXiv1202.1215.

Faircloth B.C., McCormack J.E., Crawford N.G., Harvey M.G., Brumfield R.T., Glenn T.C. 2012b. Ultraconserved elements anchor thousands of genetic markers spanning multiple evolutionary timescales. Syst. Biol. 61:717–726.

Fry B.G., Vidal N., Norman J.A., Vonk F.J., Scheib H., Ramjan S.F.R., Kuruppu S., Fung K., Hedges S.B., Richardson M.K. 2006. Early evolution of the venom system in lizards and snakes. Nature 439:584–588.

Gamble T., Coryell J., Ezaz T., Lynch J., Scantlebury D.P., Zarkower D. 2015. Restriction site-associated DNA sequencing (RAD-seq) reveals an extraordinary number of transitions among gecko sex-determining systems. Mol. Biol. Evol. 32:1296–1309.

Gatesy J., Springer M.S. 2014. Phylogenetic analysis at deep timescales: unreliable gene trees, bypassed hidden support, and the

coalescence/concatalescence conundrum. Mol. Phylogenet. Evol. 80:231–266.
Gauthier J.A., Kearney M., Maisano J.A., Rieppel O., Behlke A.D.B. 2012. Assembling the squamate tree of life: perspectives from the phenotype and the fossil record. Bull. Peabody Museum Nat. Hist. 53:3–309.
Grabherr M.G., Haas B.J., Yassour M., Levin J.Z., Thompson D.A., Amit I., Adiconis X., Fan L., Raychowdhury R., Zeng Q. 2011. Trinity: reconstructing a full-length transcriptome without a genome from RNA-Seq data. Nat. Biotechnol. 29:644–652.
Hahn, M.W. and Nakhleh, L., 2016. Irrational exuberance for resolved species trees. Evolution 70(1):7–17.
Hsiang A.Y., Field D.J., Webster T.H., Behlke A.D.B., Davis M.B., Racicot R.A., Gauthier J.A. 2015. The origin of snakes: revealing the ecology, behavior, and evolutionary history of early snakes using genomics, phenomics, and the fossil record. BMC Evol. Biol. 15:87.
Irisarri I., Baurain D., Brinkmann H., Delsuc F., Sire J.-Y., Kupfer A., Petersen J., Jarek M., Meyer A., Vences M. 2017. Phylotranscriptomic consolidation of the jawed vertebrate timetree. Nat. Ecol. Evol. 1:1370.
Jarvis E.D., Mirarab S., Aberer A.J., Li B., Houde P., Li C., Ho S.Y.W., Faircloth B.C., Nabholz B., Howard J.T. 2014. Whole-genome analyses resolve early branches in the tree of life of modern birds. Science 346:1320–1331.
Jeffroy O., Brinkmann H., Delsuc F., Philippe H. 2006. Phylogenomics: the beginning of incongruence? Trends Genet. 22:225–231.
Katoh K., Standley D.M. 2011. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol. Biol. Evol. 30:772–780.
Katzman S., Kern A.D., Bejerano G., Fewell G., Fulton L., Wilson R.K., Salama S.R., Haussler D. 2007. Human genome ultraconserved elements are ultraselected. Science 317:915.
Kearney M., Fujita M.K., Ridenour J. 2009. Lost sex in the reptiles: constraints and correlations. In: : Schön I., Martens K., van Dijk P., editors. Lost sex. London, UK: Springer. p. 447–474.
Kent W.J. 2002. BLAT—the BLAST-like alignment tool. Genome Res. 12:656–664.
Knowles L.L., Huang H., Sukumaran J., Smith S.A. 2018. A matter of phylogenetic scale: distinguishing incomplete lineage sorting from lateral gene transfer as the cause of gene tree discord in recent versus deep diversification histories. Am. J. Bot. 105:376–384.
Koch N.M., Gauthier J.A. 2018. Noise and biases in genomic data may underlie radically different hypotheses for the position of Iguania within Squamata. PLoS One 13:e0202729.
Kozlov A.M., Aberer A.J., Stamatakis A. 2015. ExaML version 3: a tool for phylogenomic analyses on supercomputers. Bioinformatics 31:2577–2579.
Kück P., Struck T.H. 2014. BaCoCa—a heuristic software tool for the parallel assessment of sequence biases in hundreds of gene and taxon partitions. Mol. Phylogenet. Evol. 70:94–98.
Lanier H.C. Huang H. Knowles L.L. 2014. How low can you go? The effects of mutation rate on the accuracy of species-tree estimation. Mol. Phylogenet. Evol. 70:112–119.
Leache A.D., Chavez A.S., Jones L.N., Grummer J.A., Gottscho A.D., Linkem C.W. 2015. Phylogenomics of phrynosomatid lizards: conflicting signals from sequence capture versus restriction site associated DNA sequencing. Genome Biol. Evol. 7:706–719.
Lemmon A.R., Emme S.A., Lemmon E.M. 2012. Anchored hybrid enrichment for massively high-throughput phylogenomics. Syst. Biol. 61:727–744.
Li H., Handsaker B., Wysoker A., Fennell T., Ruan J., Homer N., Marth G., Abecasis G., Durbin R. 2009. The sequence alignment/map format and SAMtools. Bioinformatics 25:2078–2079.
Li, H., 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv preprint arXiv:1303.3997.
Linkem C.W., Minin V.N., Leaché, A.D. 2016. Detecting the anomaly zone in species trees and evidence for a misleading signal in higher-level skink phylogeny (Squamata: Scincidae). Syst. Biol. 65: 465–477.
Losos J.B., Hillis D.M., Greene H.W. 2012. Who speaks with a forked tongue? Science 338:1428–1429.
Maddison W.P. 1997. Gene trees in species trees. Syst. Biol. 46: 523–536.
Mai U., Mirarab S. 2018. TreeShrink: fast and accurate detection of outlier long branches in collections of phylogenetic trees. BMC Genomics 19:272.
Nute M., Chou J., Molloy E.K., Warnow T. 2018. The performance of coalescent-based species tree estimation methods under models of missing data. BMC Genomics 19:286.
Paradis E., Claude J., Strimmer K. 2004. APE: analyses of phylogenetics and evolution in R language. Bioinformatics 20:289–290.
Philippe H., Brinkmann H., Lavrov D. V, Littlewood D.T.J., Manuel M., Wörheide G., Baurain D. 2011. Resolving difficult phylogenetic questions: why more sequences are not enough. PLoS Biol. 9:e1000602.
Pisani D., Pett W., Dohrmann M., Feuda R., Rota-Stabelli O., Philippe H., Lartillot N., Wörheide G. 2015. Genomic data do not support comb jellies as the sister group to all other animals. Proc. Natl. Acad. Sci. USA 112:15402–15407.
Prum R.O., Berv J.S., Dornburg A., Field D.J., Townsend J.P., Lemmon E.M., Lemmon A.R. 2015. A comprehensive phylogeny of birds (Aves) using targeted next-generation DNA sequencing. Nature 526:569–573.
Pyron R.A., Burbrink F.T. 2014. Early origin of viviparity and multiple reversions to oviparity in squamate reptiles. Ecol. Lett. 17:13–21.
Pyron R.A., Burbrink F.T., Wiens J.J. 2013. A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. BMC Evol. Biol. 13:93.
Reddy S., Kimball R.T., Pandey A., Hosner P.A., Braun M.J., Hackett S.J., Han K.-L., Harshman J., Huddleston C.J., Kingston S. 2017. Why do phylogenomic data sets yield conflicting trees? Data type influences the avian tree of life more than taxon sampling. Syst. Biol. 66:857–879.
Reeder T.W., Townsend T.M., Mulcahy D.G., Noonan B.P., Wood Jr P.L., Sites Jr J.W., Wiens J.J. 2015. Integrated analyses resolve conflicts over squamate reptile phylogeny and reveal unexpected placements for fossil taxa. PLoS One 10:e0118199.
Revell L.J. 2012. phytools: an R package for phylogenetic comparative biology (and other things). Methods Ecol. Evol. 3:217–223.
Rokas A., Carroll S.B. 2006. Bushes in the tree of life. PLoS Biol. 4:e352.
Romiguier, J., Cameron, S.A., Woodard, S.H., Fischman, B.J., Keller, L. and Praz, C.J., 2016. Phylogenomics controlling for base compositional bias reveals a single origin of eusociality in corbiculate bees. Mol. Biol. Evol. 33: 670–678.
Salichos L., Rokas A. 2013. Inferring ancient divergences requires genes with strong phylogenetic signals. Nature 497:327–331.
Schliep K.P. 2010. phangorn: phylogenetic analysis in R. Bioinformatics 27:592–593.
Shen X.-X., Hittinger C.T., Rokas A. 2017. Contentious relationships in phylogenomic studies can be driven by a handful of genes. Nat. Ecol. Evol. 1:126.
Singhal S., Grundler M., Colli G., Rabosky D.L. 2017. Squamate conserved loci (Sq CL): a unified set of conserved loci for phylogenomics and population genetics of squamate reptiles. Mol. Ecol. Resour. 17:e12–e24.
Smith S.A., Moore M.J., Brown J.W., Yang Y. 2015. Analysis of phylogenomic datasets reveals conflict, concordance, and gene duplications with examples from animals and plants. BMC Evol. Biol. 15:150.
Smith S.A., O'Meara B.C. 2012. treePL: divergence time estimation using penalized likelihood for large phylogenies. Bioinformatics 28:2689–2690.
Smith S.A., Walker J.F., Brown J., Walker-Hale N. 2020. Nested phylogenetic conflicts, combinability, and deep phylogenomics in plants. Syst. Biol. 69:579–592.
Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30:1312–1313.
Streicher J.W., Schulte J.A., Wiens, J.J., 2016. How should genes and taxa be sampled for phylogenomic analyses with missing data? An empirical study in iguanian lizards. Syst. Biol. 65:128–145.
Streicher J.W., Wiens J.J. 2016. Phylogenomic analyses reveal novel relationships among snake families. Mol. Phylogenet. Evol. 100:160–169.

Streicher J.W., Wiens J.J. 2017. Phylogenomic analyses of more than 4000 nuclear loci resolve the origin of snakes among lizard families. Biol. Lett. 13:20170393.

Tonini J.F.R., Beard K.H., Ferreira R.B., Jetz W., Pyron R.A. 2016. Fully-sampled phylogenies of squamates reveal evolutionary patterns in threat status. Biol. Conserv. 204:23–31.

Townsend J.P. 2007. Profiling phylogenetic informativeness. Syst. Biol. 56:222–231.

Townsend T.M., Larson A., Louis E., Macey J.R. 2004. Molecular phylogenetics of Squamata: the position of snakes, amphisbaenians, and dibamids, and the root of the squamate tree. Syst. Biol. 53:735–757.

Uetz P., Stylianou A. 2018. The original descriptions of reptiles and their subspecies. Zootaxa 4375:257–264.

Van der Auwera, G.A., Carneiro, M.O., Hartl, C., Poplin, R., Del Angel, G., Levy-Moonshine, A., Jordan, T., Shakir, K., Roazen, D., Thibault, J. and Banks, E. 2013. From FastQ data to high-confidence variant calls: the genome analysis toolkit best practices pipeline. Curr. Protoc. Bioinformatics 43:11.

Walker J.F., Brown J.W., Smith S.A. 2018. Analyzing contentious relationships and outlier genes in phylogenomics. Syst. Biol. 67:916–924.

Wen D., Yu Y., Zhu J., Nakhleh L. 2018. Inferring phylogenetic networks using PhyloNet. Syst. Biol. 67:735–740.

Whelan N.V, Kocot K.M., Moroz L.L., Halanych K.M. 2015. Error, signal, and the placement of Ctenophora sister to all other animals. Proc. Natl. Acad. Sci. USA 112:5773–5778.

Whelan N. V, Kocot K.M., Moroz T.P., Mukherjee K., Williams P., Paulay G., Moroz L.L., Halanych K.M. 2017. Ctenophore relationships and their placement as the sister group to all other animals. Nat. Ecol. Evol. 1:1737.

Wiens J.J., Hutter C.R., Mulcahy D.G., Noonan B.P., Townsend T.M., Sites Jr J.W., Reeder T.W. 2012. Resolving the phylogeny of lizards and snakes (Squamata) with extensive sampling of genes and species. Biol. Lett. 8:1043–1046.

Wilke C.O. 2016. Cowplot: streamlined plot theme and plot annotations for 'ggplot2'. 2016. URL https//CRAN. R-project. org/package= cowplot. R Packag. version 0.7. 0. p. 287.

Wong K.M., Suchard M.A., Huelsenbeck J.P. 2008. Alignment uncertainty and genomic analysis. Science 319:473–476.

Zhang C., Rabiee M., Sayyari E., Mirarab S. 2018. ASTRAL-III: polynomial time species tree reconstruction from partially resolved gene trees. BMC Bioinformatics 19:153.

Zhang J., Kobert K., Flouri T., Stamatakis A. 2013. PEAR: a fast and accurate Illumina Paired-End reAd mergeR. Bioinformatics 30:614–620.